

Noise-induced rhythmicity in an ensemble of circadian oscillators



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- Acknowledgement

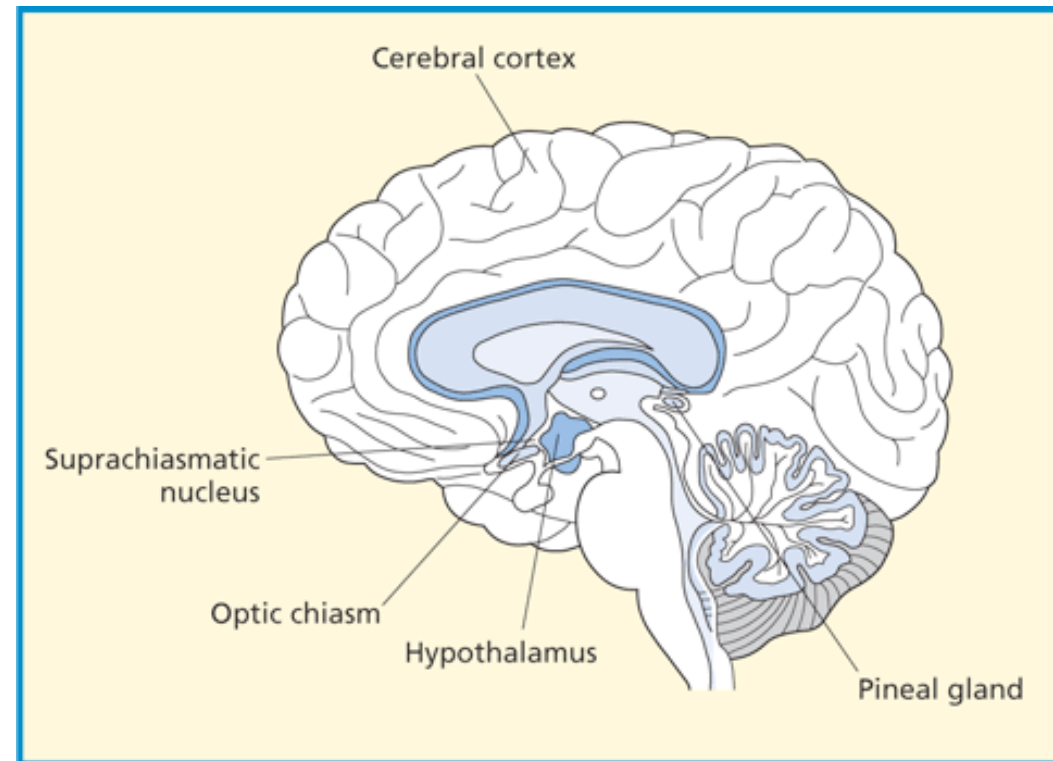
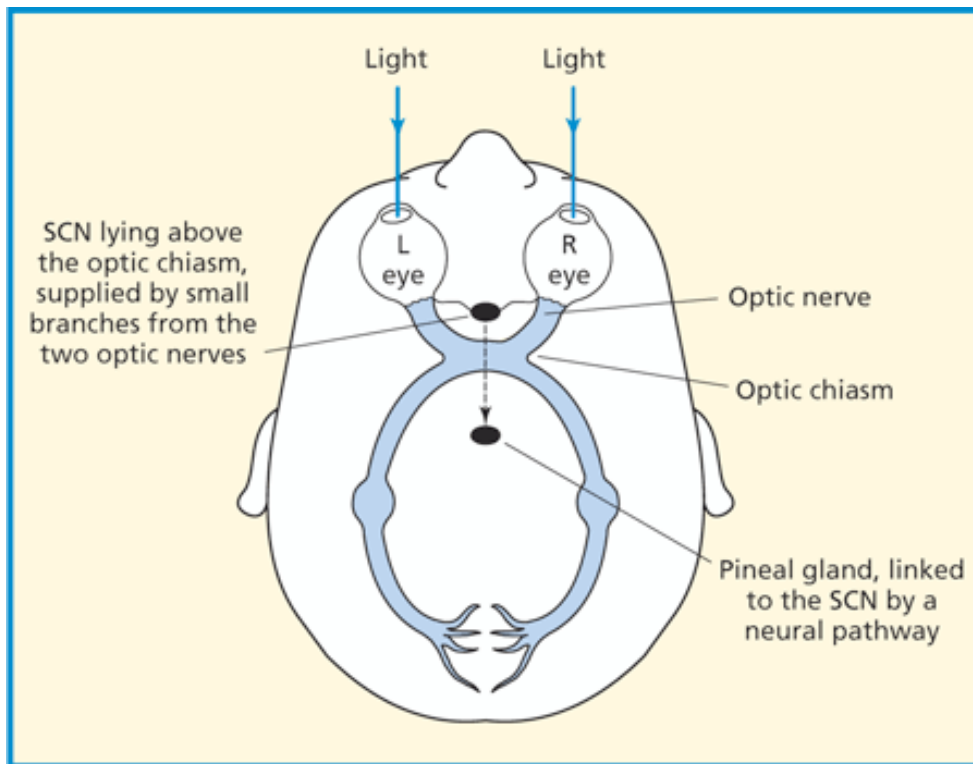
The normal Circadian Rhythm

a roughly-24-hour cycle in the biochemical, physiological or behavioral processes of living beings



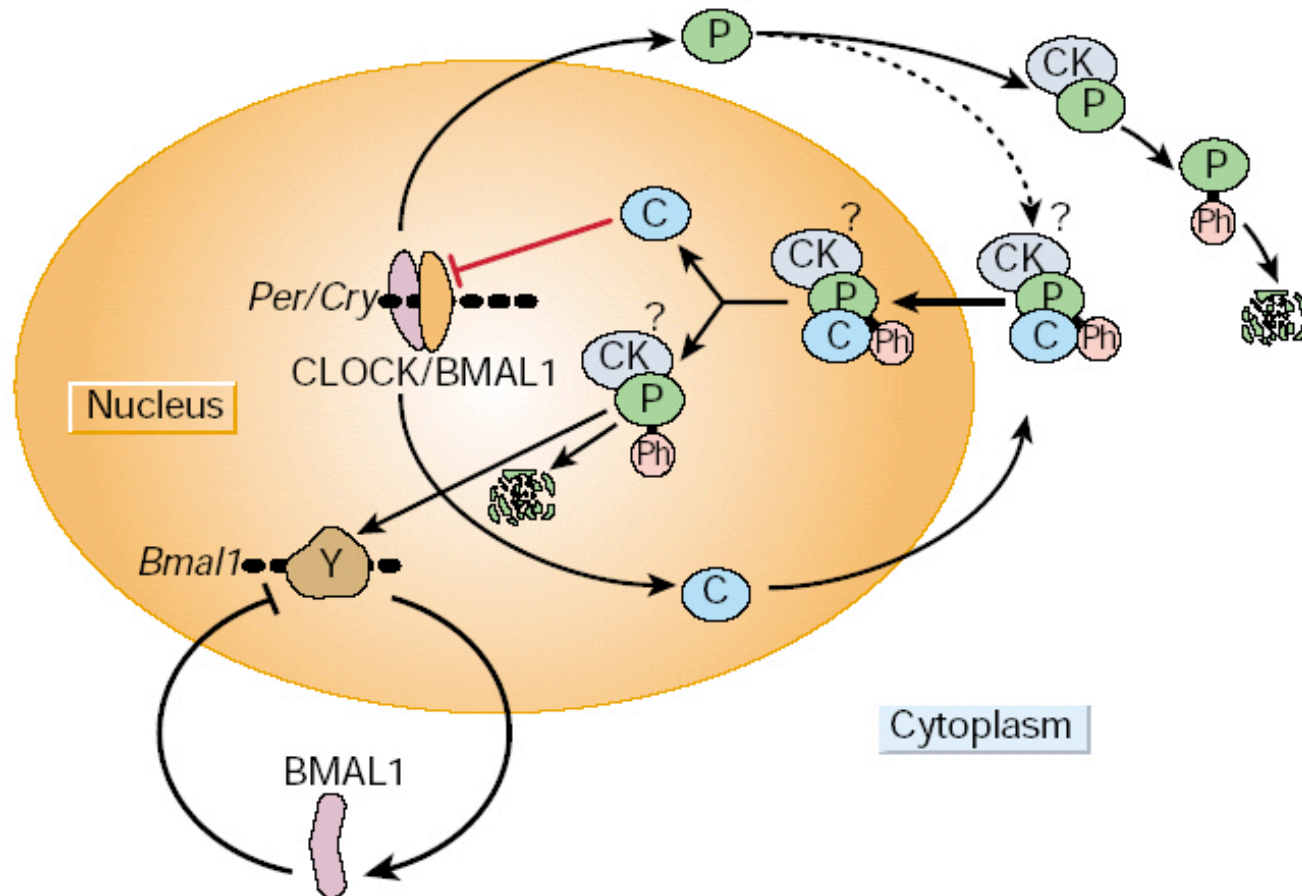
The circadian pacemaker

The Suprachiasmatic Nucleus



The circadian pacemaker

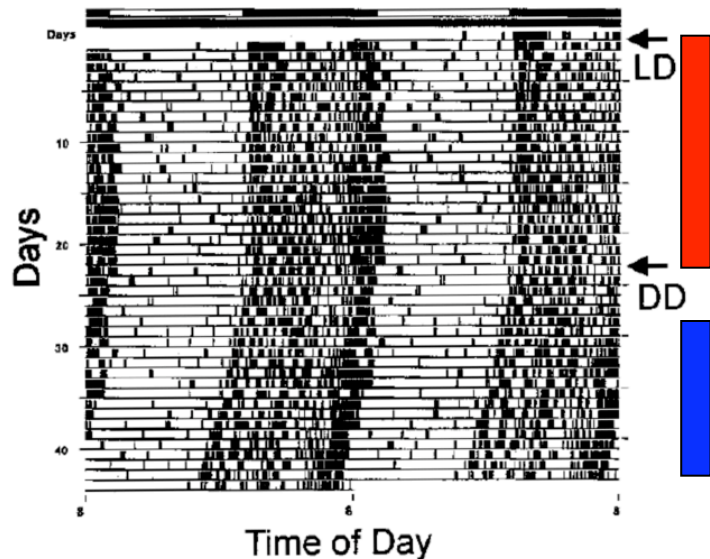
Mammalian circadian clock



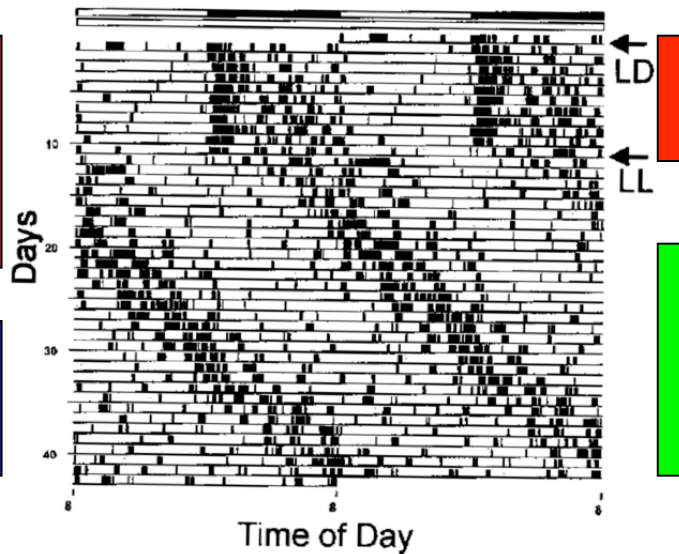
S. Panda, J.B. Hogenesch and S.A. Kay,
Nature, 2002, **417**, 329-335

The transitions

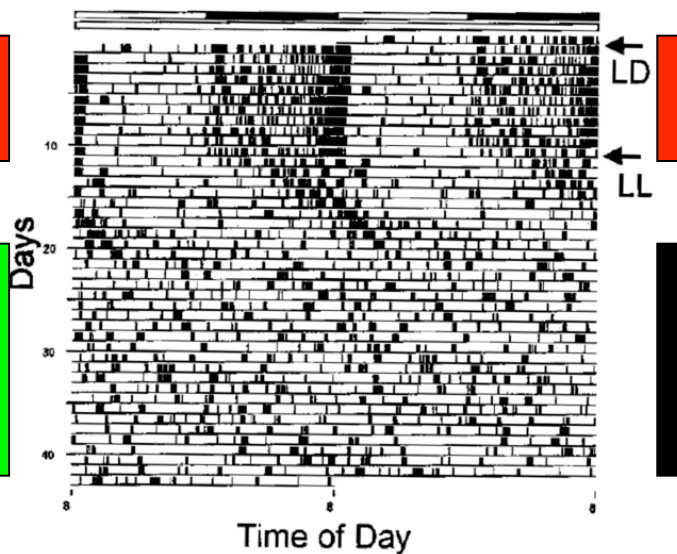
LD → DD







LD → LL



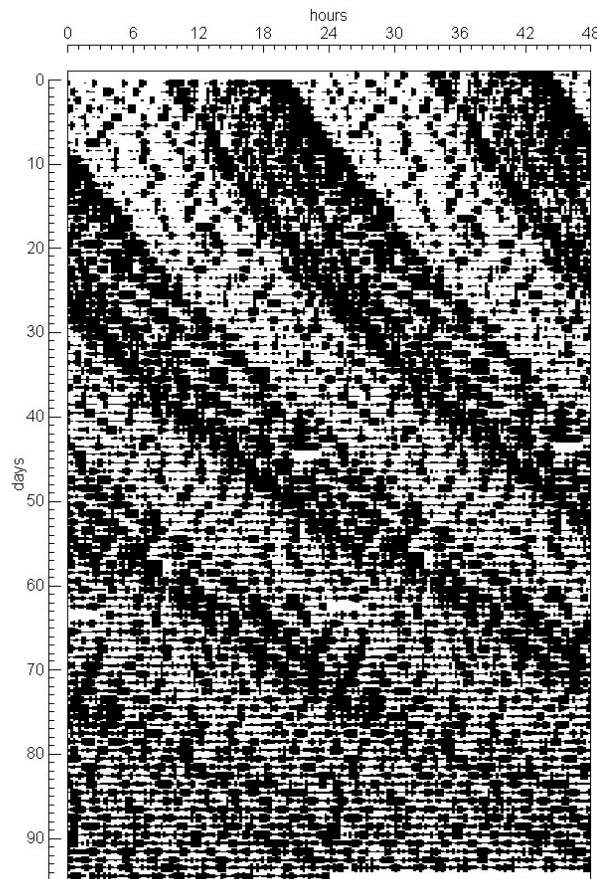
LD → LL



-  rhythmic entrain
-  rhythmic, free running, $T < 24$ h
-  rhythmic, free running, $T > 24$ h
-  arrhythmic

The transitions in experiments

stepwise light increase



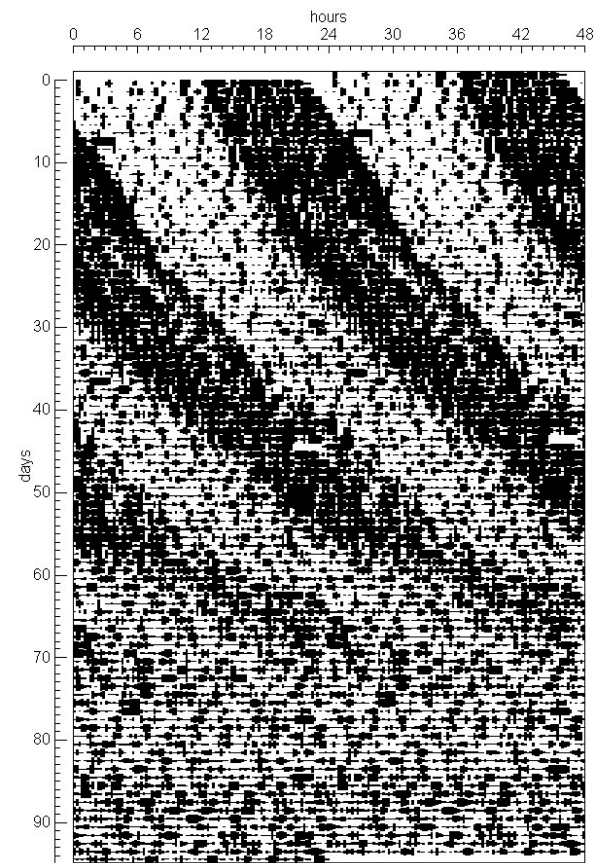
0.02 lux

0.14 lux

1 lux

6 lux

300 lux



The transition on the genetic level

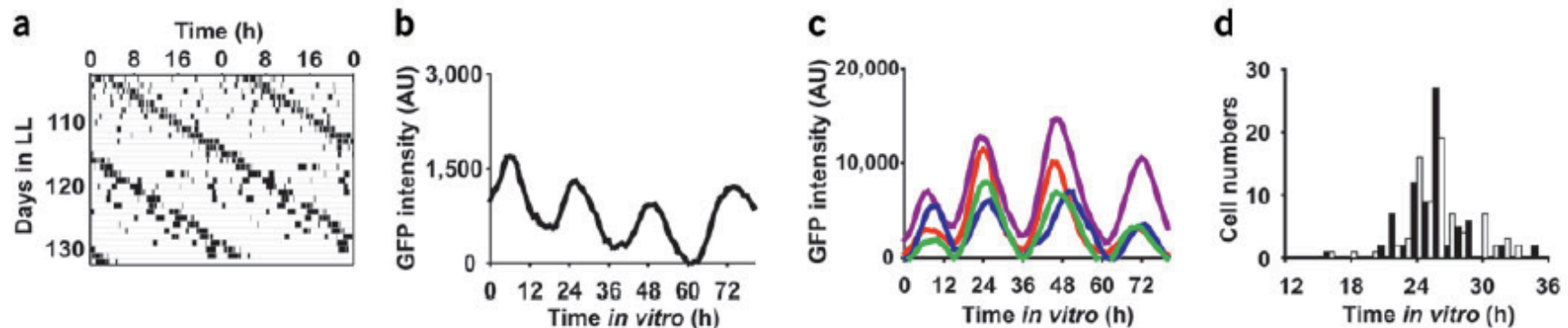


Figure 2 Behavioral and SCN rhythms from a rhythmic constant light-treated mouse. (a) Actogram of wheel running activity from a mouse that remained rhythmic in LL. (b) Time-lapse SCN *Per1:GFP* fluorescence signals. (c) Individual SCN neuronal *Per1:GFP* rhythms from SCN in b. (d) Peak time histograms of individual neuronal rhythms. $n = 154$ cells.

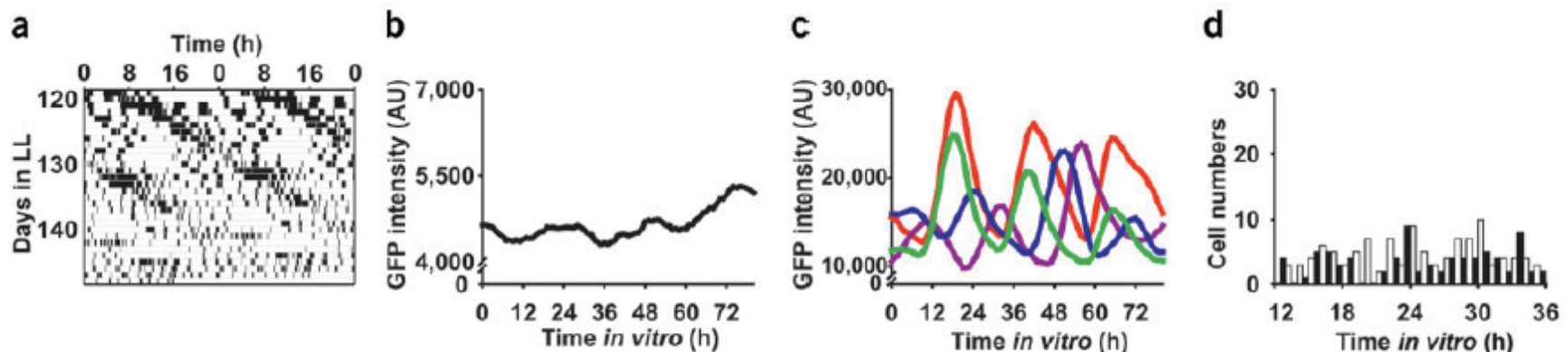


Figure 1 Behavioral and SCN rhythms from an arrhythmic constant light-treated mouse. (a) Actogram of wheel running activity. Black marks indicate wheel revolutions. Note loss of temporal organization in latter portion of record. (b) Time-lapse SCN *Per1:GFP* fluorescence signals for 3.5 d *in vitro*. (c) Individual SCN neuronal *Per1:GFP* rhythms from SCN in b. Four representative cells are plotted for clarity (colored lines). (d) Peak time histograms of individual neuronal rhythms. Peak times of neurons in the right SCN are plotted with black bars, whereas those in the left SCN are plotted with open bars. Histograms for this and the following figures are for hours 12–36 *in vitro*. $n = 193$ cells. Animal care and use was reviewed and approved by the Vanderbilt University IACUC.

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- Each genetic oscillator is self-oscillatory
- Light influences the coupling
- The circadian rhythm is a joint effect of all oscillators

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- Can noise restore a rhythm in the arrhythmic state?

The modified Goodwin model

$$\begin{aligned}\frac{dX_i}{dt} &= \nu_1 \frac{K_1^n}{K_1^n + Z_i^n} - \nu_2 \frac{X_i}{K_2 + X_i} + \nu_c \frac{KV_i}{K_c + KV_i} && \text{clock gene} \\ \frac{dY_i}{dt} &= k_3 X_i - \nu_4 \frac{Y_i}{K_4 + Y_i} && \text{clock protein} \\ \frac{dZ_i}{dt} &= k_5 Y_i - \nu_6 \frac{Z_i}{K_6 + Z_i} && \text{active protein} \\ \frac{dV_i}{dt} &= k_7 X_i - \nu_8 \frac{V_i}{K_8 + V_i} - \eta(V_i - (Q_0 + \zeta(t))F) && \text{neurotransmitter} \\ F &= \frac{1}{N} \sum_{i=1}^N V_i\end{aligned}$$

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 \end{aligned}$$

$\nu_{1,2,4,6,8}$ and $k_{3,5,7}$ individually rescaled by τ_i

τ_i normal distributed with diversity σ_τ

→ individual eigen-frequencies

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$$R_{syn} = \frac{\langle \bar{Y}^2 \rangle - \langle \bar{Y} \rangle^2}{\frac{1}{N} \sum_{i=1}^N (\langle Y_i^2 \rangle - \langle Y_i \rangle^2)} = \frac{Var(\bar{Y})}{Mean_i(Var_t(Y_i))}$$
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$R_{syn}=1 \rightarrow$ complete synchronized

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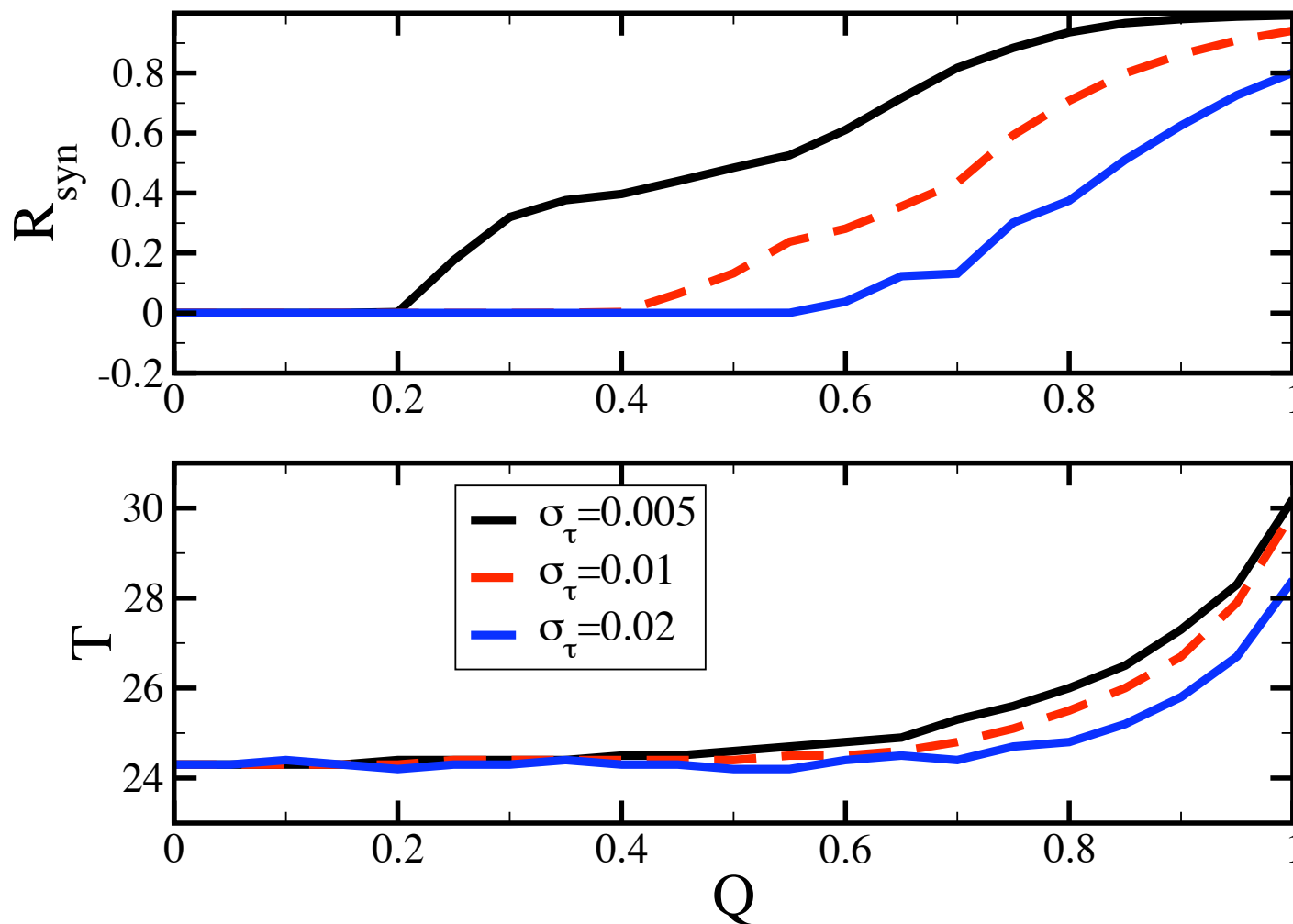
- The **coherence** measure:

the decay time of the envelope of the autocorrelation function of the overt rhythm

The transition

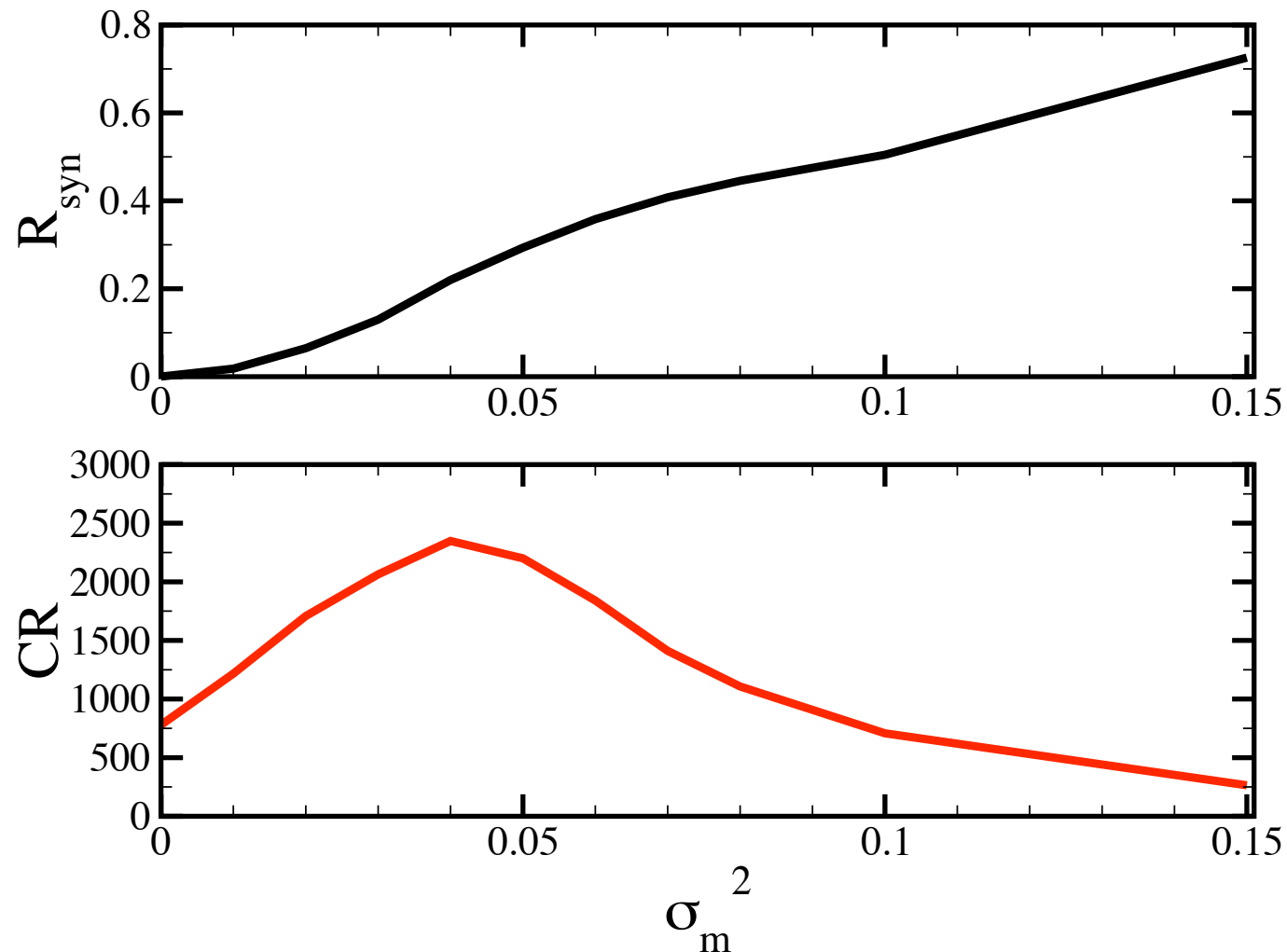
arrhythmic \leftrightarrow rhythmic

unsynchronized \leftrightarrow synchronized



The Coherence Resonance Noise-induced rhythmicity

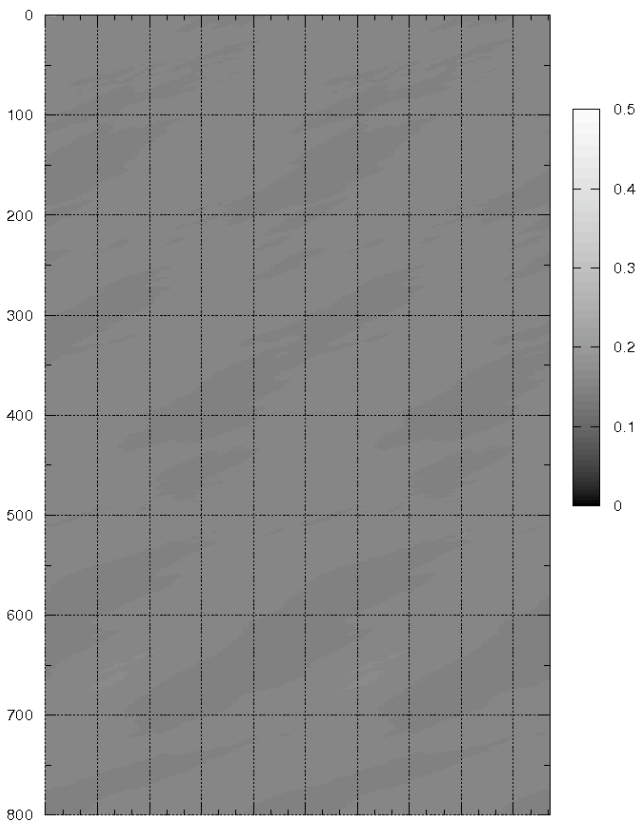
$$\sigma_\tau = 0.005, Q_0 = 0.15$$



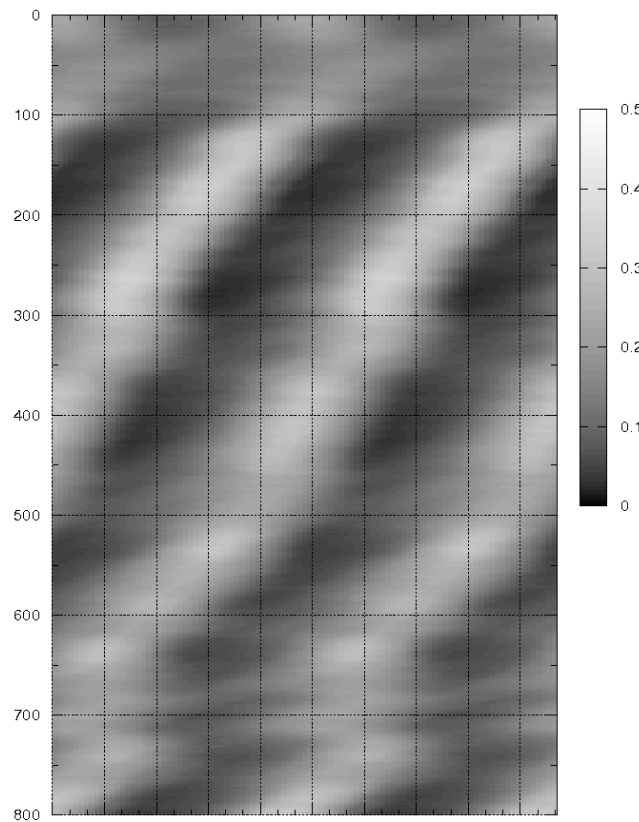
CR in Double plotted actograms

$$\sigma_{\tau} = 0.005, Q_0 = 0.15$$

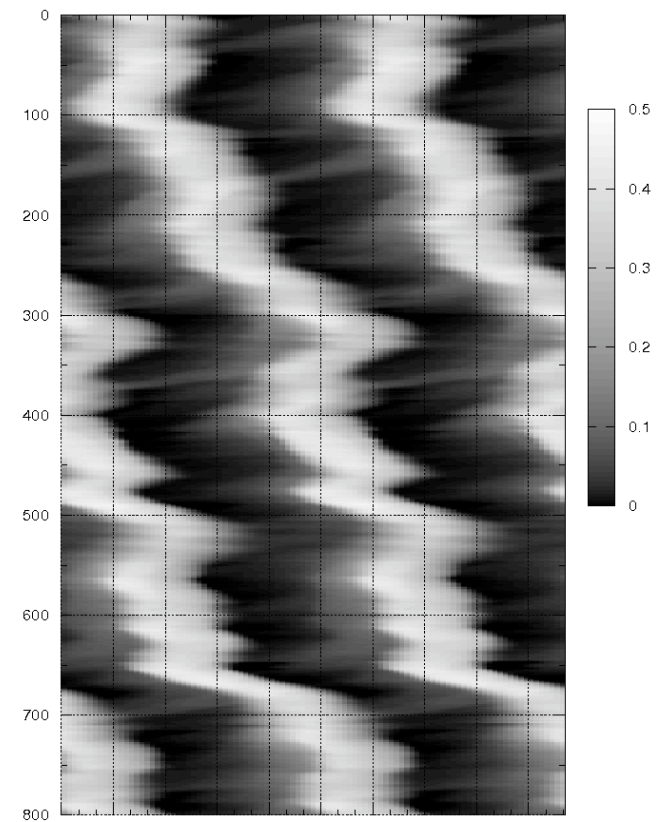
$$\sigma_m^2 = 0.0$$



$$\sigma_m^2 = 0.04$$

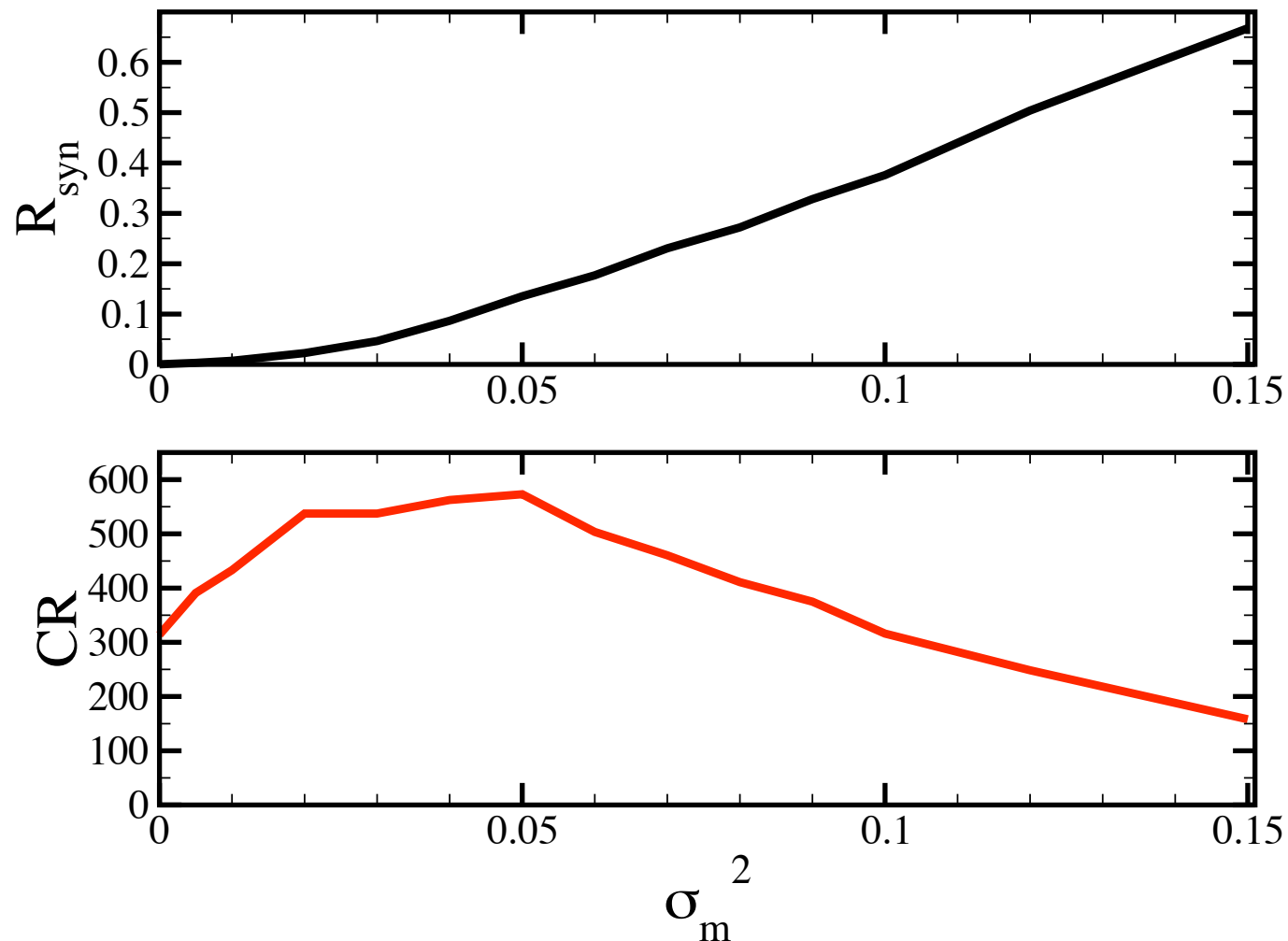


$$\sigma_m^2 = 0.1$$



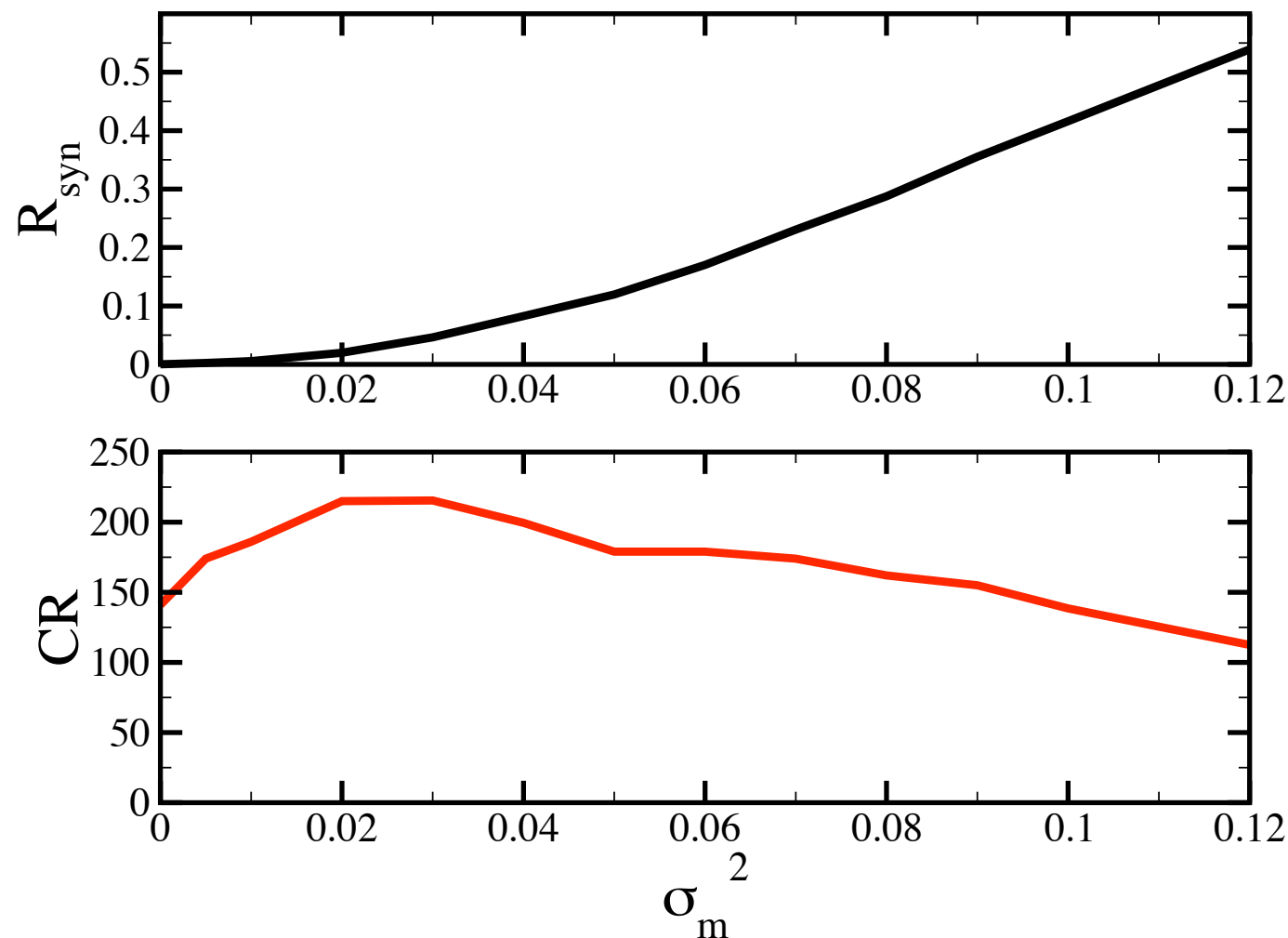
The Coherence Resonance Noise-induced rhythmicity

$$\sigma_\tau = 0.01, Q_0 = 0.25$$



The Coherence Resonance Noise-induced rhythmicity

$$\sigma_\tau = 0.02, Q_0 = 0.4$$



Conclusion

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- Noise is multiplicative and affects the coupling strength

Open Problem on Noise

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- The hypothesis of light-dependent coupling is critical for CR in the model
- Would a hypothetical noise-enhanced rhythmicity in *in vivo* experiments be sufficient to strengthen the hypothesis of light-dependent coupling?

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**Thank you for your
attention and interest**